This is an open-access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)



Review Article

https://doi.org/10.47278/journal.ijvs/2023.108

Role of Programmed Cell Death Receptor-1 and Cytotoxic T Lymphocyte-Associated Antigen 4 in Bovine Leukemia Virus Infection

Kassym Mukanov, Kanatbek Mukantayev and Kanat Tursunov*

National Center for Biotechnology, Ministry of Healthcare of the Republic of Kazakhstan, 010000, Astana, Kazakhstan ***Corresponding author:** tursunov@biocenter.kz

ABSTRACT

A cluster of T-cell receptors includes activating and inhibitory stimulatory molecules that favorably or unfavorably control immune responses. Recent studies on chronic bovine infections have revealed that, under severe viral loads and malignant pathologies, stromal and immune cells increase the expression of immune inhibitory molecules. To maintain internal homeostasis, programmed cell death receptor-1 (PD-1) and cytotoxic T lymphocyte-associated antigen-4 (CTLA-4) inhibit T cell activity. In chronic viral infections, the prolonged activation of T cells leads to the continuous production of PD-1 and CTLA-4. Blocking PD-1 and CTLA-4 is a successful therapeutic approach that is actively used in the treatment of oncological diseases. The effectiveness of this approach in the treatment of chronic viral infections, particularly those caused by bovine leukemia virus has been hypothesized. However, uncertainty surrounds these receptors' function in persistent viral infections. This review focused on the functions of PD-1 and CTLA-4 in bovine leukemia virus and discusses disease therapies based on their blockade.

Key words: Programmed cell death receptor-1, Cytotoxic T lymphocyte-associated antigen 4, T cell exhaustion, Chronic virology infection, Checkpoint inhibitor, Bovine leukemia virus.

INTRODUCTION

The oncogenic Bovine Leukemia Virus (BLV) belongs to the genus Deltaretrovirus, family Retroviridae, and subfamily Orthoretrovirinae. The virus is widespread in domestic cattle worldwide, affecting up to 39% of beef breeds and up to100% of dairy breeds. Despite the ability of the virus to infect blood and milk, less than 5% of infected cattle show clinical signs of the disease. The mechanism of BLV transmission to humans is unknown; however, consumption of raw milk can transmit the virus from cattle to humans (Buehring et al. 2019; Khatami et al. 2020; Canova et al. 2021; de Quadros et al. 2023).

Despite ongoing anti-BLV interventions, the widespread prevalence of the disease has increased interest in studying immune checkpoints for the treatment of chronic bovine infections. The programmed cell death receptor-1 (PD-1)/programmed cell death ligand (PD-L1) signaling pathway is associated with BLV infection. Studies have shown that immune suppressive molecules are highly expressed as the BLV infection progresses (Shirai et al. 2011; do Nascimento et al. 2023). The regulatory mechanisms of the immune system under

different physiological conditions are based on the opposing activities of various T helper cell subpopulations. T cells are one of the critical cells that protect the organism from pathogenic microbes, maintain tolerance, and reduce tumor progression and metastasis (Jubel et al. 2020; Zou and Chen 2008). Effectors T cells (Teffs), which include regulatory T cells (Tregs), helper T cells (Ths), and cytotoxic T cells (CTL), mediate the antagonistic activity of the T-helper cell subpopulation. The roles of Teffs in acquiring immunity and Tregs in developing tolerance are essential for maintaining effective immunity and internal homeostasis (Bucktrout et al. 2018).

The opposing activities of Teffs and Tregs are regulated by several receptors that activate or inhibit signals. Stimulatory or inhibitory receptor signals are activated upon binding of the T-cell receptor (TCR) to the major histocompatibility complex (MHC). These stimulatory and inhibitory signaling mechanisms provide additional information to T cells concerning local microenvironment and host state (Frauwirth et al. 2002; Parry et al. 2005). One of the receptors regulating T-cell activity is PD-1 and its ligands, PD-L1 and PD-L2, which play essential roles in both the T-cell activity regulated

Cite This Article as: Mukanov K, Mukantayev K and Tursunov K, 2024. Role of programmed cell death receptor-1 and cytotoxic T lymphocyte-associated antigen 4 in bovine leukemia virus infection. International Journal of Veterinary Science 13(3): 369-377. <u>https://doi.org/10.47278/journal.ijvs/2023.108</u>

negatively and positively. The PD-1 protein reduces T lymphocytes activation, thereby reducing the risk of autoimmunity and immunopathology (Freeman et al. 2000; Latchman et al. 2001; Sharpe and Pauken 2018). Further, the B7 family receptor co-stimulator, cytotoxic T lymphocyte-associated antigen 4 (CTLA-4), exert inhibitory and stimulatory effects on activation. The CTLA-4 receptor is a competitor of the cluster of differentiation (CD28) receptor, thereby inhibiting the formation of the CD28:B7 signaling pathway and reducing immune activation. In addition, the CTLA-4 receptor can disrupt stimulatory TCR and MHC signaling and bind to the B7.1 receptor CD80 on dendritic cells to inhibit antitumor immunity (Buchbinder and Desai 2016; Shen and Zhao 2018; Mayoux et al. 2020). In addition to CTLA-4's ability to inhibit the immune system, increased receptor expression has been observed during the progression of human immunodeficiency virus (HIV) infection. High receptor expression levels have also been observed in liver CD8+ T cells during viral hepatitis (Nakamoto et al. 2009; Zhang et al. 2010). A worse prognosis is linked to increased CTLA-4 expression in nasopharyngeal cancer patients (Zhang et al. 2016). The presented data demonstrates the role of CTLA-4 receptor in aggravating not only cancer but also chronic infections. In addition, dysfunction of antigen-presenting cells (APCs), cells of myeloid origin, Tregs, and stromal and tumor cells with high levels of CTLA-4 expression are essential factors for decreased immunity against chronic infections and cancer (Chen 2004; Curiel et al. 2004; Gabrilovich 2004; Banchereau and Palucka 2005).

Given the regulatory functions of PD-1 and CTLA-4 receptors in inflammatory processes and tumors, chronic infectious agents and transformed cells have evolved mechanisms to evade host immunity (Attanasio and Wherry 2016; LaFleur et al. 2018). Thus, the study of PD-1 and CTLA-4 receptor function during chronic infection or in oncology is an area of intense research. This review presents the structures of PD-1 and CTLA-4 receptors and their functional roles in bovine leukemia. Based on the literature, the therapeutic roles of PD-1 and CTLA-4 receptor blockade involving different mechanisms of immune inhibition in BLVinfected cows have been described.

Structure and Function of PD-1

Under typical physiological conditions, the body uses the PD-1 signaling pathway to induce apoptosis to limit excessive T cell activation in peripheral organs. The same signaling mechanism controls the immune response to bacterial and viral infections. The inhibition of PD-L1 and PD-1 receptors restored cytotoxic T-cell growth and cytokine expression in CD4-deficient and infected virus mice. By eliminating infected cells, cytotoxic T cells can lower the viral burden (Barber et al. 2006; Francisco et al. 2010). Blocking the PD-1/PD-L1 or PD-1/PD-L2 signaling pathways leads to a similar effect in cancer (Dong et al. 2016; Rui et al. 2023) (Fig. 1).

The PD-1 receptor CD279 was isolated from hybrid cells of murine T cells and a progenitor hematopoietic cell line. On chromosome 2 (2q37), the conserved regions (CR)-B and CR-C of the programmed cell death protein 1 (Pdcd1) gene, which codes for the PD-1 receptor, are two Deoxyribonuclease I hypersensitive sites that affect

receptor expression. Nuclear factor activating T cell transcription (NFAT) is present in CR-C and is essential in Pdcd1 expression. In addition, NFATc1 binding to CR-C and c-Fos sites in the CR-B region of CD4+ and CD8+ T cells enhances PD-1 production at the initial recognition stage. The receptor is a membrane protein refers to the CD28 family. At the protein's extracellular N terminus, there is an IgV-like fragment, a transmembrane fragment, and a cytoplasmic fragment. The receptor has a molecular weight of 55 kDa and a length of 288 aa. There are two amino acid sequences in the cytoplasmic domain of PD-1. that is, tyrosine-based inhibitory and switch motifs. Tyrosine phosphatases (SHP) 1 and SHP-2, which contain the sarcoma homology 2 domain, are connected to the Cterminal tyrosine sequence (TEYATIVF). Protein tyrosine phosphorylation and dephosphorylation are critical regulatory activities in numerous signaling pathways that result in cell growth, differentiation, and death (Ishida et al. 1992; Shinohara et al. 1994; Starr et al. 1997; Lorenz 2009).

Immune cells that have been activated, such as CD4+ T cells, CD8+ T cells, B cells, T-killer cells, monocytes, dendritic cells, and macrophages, express the Pdcd1 gene. Additionally, Pdcd1 expression is an indicator for eliminated T lymphocytes and cells with reduced effector function and is specifically elevated in T cells exposed to long-term antigens (Agata et al. 1996; Matsuzaki et al. 2010).

The activation of the PD-1 receptor is regulated by several mechanisms. PD-L1 production in cells of tumor and increased PD-1 production in CD8+ T-lymphocytes correlate with soluble factors like interleukins 6 and 10 (IL6, IL-10) (Chen 2004: Curiel et al. 2004). Additionally, there was an indicated association between the production of PD-L1 on monocytes in the blood and that of PD-1 on circulating CD4+ or CD8+ T cells, suggesting that the same mechanisms may be responsible for the increased activity of PD-1 and PD-L1. Furthermore, the production of PD-1 in both CD4+ and CD8+ T lymphocytes was shown to be significantly higher inside tumor tissues when in contrast to cells from samples of blood and healthy stomach mucosa. The findings imply that in gastric tumor, cancer cells influence the production of PD-1 and PD-L1 (Saito et al. 2013).

The transcription factor T-box protein produced by T cells (T-bet) also controls PD-1 expression. An investigation on the stimulation of killer T cells against the virus in persistent infections shed light on the function of T-bet. Depending on the level of immune activity during chronic viral infections, the relationship between PD-1 and T-bet protein may also change. For example, during 15 days of viral infection, no relationship was found between the levels of PD-1 receptor and T-bet protein. Additionally, decreased T-bet expression during acute infection leads to increased PD-1 production in CD8+ T cells. However, for full production of PD-1 at persistent infections, receptor regulation by T-bet is insufficient. Antigenic signals, T-bet, and other transcription factors are expected to control PD-1 and other regulatory receptors (Kao et al. 2011).

The two major PD-1 ligands are PD-L1 and PD-L2. The ligand of PD-1 is a 290 aa long membrane-spanning glycoprotein member of the Ig superfamily B7-CD28 (Kao et al. 2011). The B7 family of proteins includes B7-DC,

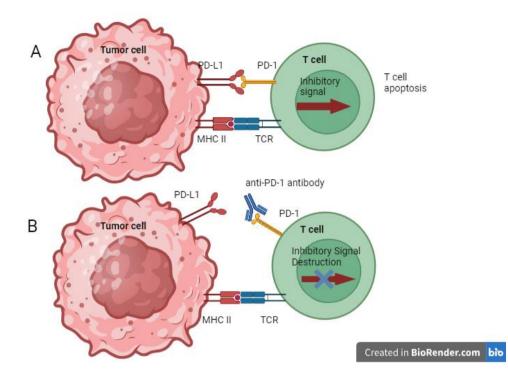


Fig. 1: PD-1-mediated inhibition of T cells. A. Tumor cells evade the immune response by activating PD-1 using PD-L1 ligand. B. Binding of anti-PD-1 antibodies to PD-1 increases T cell activity against tumorigenic cells.

also known as PD-L2, which is the second recognized ligand of the PD-1 receptor. In all healthy human and mouse tissues, PD-L1 messenger ribonucleic acid (mRNA) expression was found. However, it has been noted that the PD-L1 receptor is present on the surface of some macrophage-like cells in internal organs. The disparity between the mRNA and receptor expression on the cell membrane highlights the crucial function of the posttranscriptional systems regulating PD-L1 production (Sanmamed and Chen 2014).

Structure and Function of CTLA-4

CTLA-4 or CD152 belongs to the Ig superfamily and has been identified as a membrane protein with a molecular mass of approximately 41–43 kDa. CTLA-4 generally consists of a leader peptide, extracellular, membrane and intracellular region. Three protein isoforms have been identified as a result of standard and alternative CTLA-4 mRNA splicing. The surface CTLA-4 protein is the first, the second is soluble CTLA-4 (sCTLA-4) protein with the transmembrane fragment removed, and the third is the CTLA-4 independent from ligand (liCTLA-4) protein lacking the extracellular domain (Jakubczik et al. 2016).

The presence of the receptor on the outer layer of T cells during the G1 stage and subsequent phases of the cell lifecycle is suggested by the early occurrence of CTLA-4 mRNA expression during the stimulation of T cells. A lack of protein mRNA expression leads to severe autoimmune diseases and significant tissue damage in many organs (Sutherland et al. 2000; Homann et al. 2006). Various nucleotide substitutions in CTLA-4 gene mapped to human chromosome 2q33 have been associated with susceptibility to various autoimmune and infectious diseases mediated by T cells (Danilovic et al. 2012; Liu et al. 2013). According to Eskandari-Nasab et al. (2014), the CTLA-4 gene C/T polymorphism at position 318 in the Iranian population

indicated susceptibility to the risk of brucellosis infection. The leader peptide codon is altered by the CTLA-4 A/G polymorphism at position +49, which causes alanine to be changed to threonine. There are no statistically significant variations between autoimmune illness patients and healthy people in the frequencies of the alleles and genotypes of the CTLA-4 gene variations, according to studies done in patients with autoimmune diseases. For example, CTLA-4 gene polymorphism was not associated with systemic lupus erythematosus (Farivar et al. 2014; Oaks and Hallett 2000; Rochmah et al. 2022).

Activated T cells produce the CTLA-4 receptor, which interacts with APCs via B7-1 and B7-2 (Lindsten et al. 1993; Mulley and Nikolic-Paterson 2008; Wing et al. 2011) (Fig. 2). This receptor was the first immune checkpoint that has become a clinical target for cancer immunotherapy. Monoclonal antibodies against CTLA-4 blocking the CTLA-4-CD80/CD86 signaling pathway activate antitumor immunity and improve the survival of patients with melanoma (Danilovic et al. 2012). One signaling pathway that enhances T-cell activity is the interaction of the CD28 receptor with CD80 and CD86, which significantly enhances the TCR-antigen signaling. In contrast to CD28, CTLA-4 has stronger affinity to CD80 and CD86, and its induction negatively regulates T-cell activation (Freeman et al. 1993; Hathcock et al. 1993; Linsley et al. 1994: Lee et al. 1998: Rudd et al. 2009: Schneider et al. 2006; Fife and Bluestone 2008; Callahan et al. 2010).

The B7-1 and B7-2 receptors play a crucial role in activating T cells by transmitting a signal from CD28 upon specific binding of the TCR receptor to an antigen on the MHC. B7-1 and B7-2 are transmembrane proteins type 1 that have the proximal IgC and distal IgV membrane domains, respectively. Although both proteins interact with CTLA-4 and CD28, differences exist in the strength of the

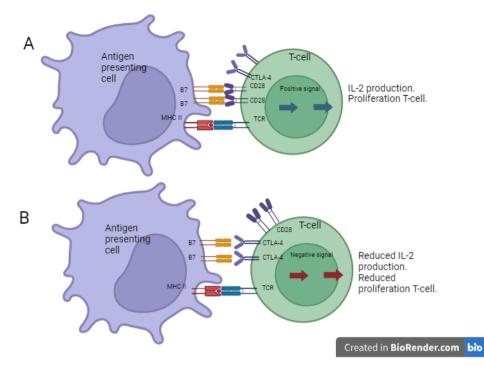


Fig. 2: Inhibition of T lymphocytes by CTLA-4 protein. A. Under weak T-cell receptor (TCR) stimulation, CD28 and B7 receptors are predominantly dissociated, which is a positive signal for interleukin 2 (IL2) productions and T-lymphocytes proliferation. B. Under intense TCR stimulation, there is an increase in CTLA-4 induction and dissociation of CTLA-4 and CD28 receptors, which results in the inhibition of IL-2 and T-cells.

equilibrium dissociation constants between these receptors. The equilibrium dissociation constant of B7-1 with CTLA-4 and CD28 was 5–10 times greater than that of B7-2. The structure of receptors' cytoplasmic tails and the molecules engaged in the signaling pathway are probably related to the differences in the biological functions that exist amongst receptors. B7-1 and B7-2 receptors' extracellular domains' monomeric or dimeric states play a significant role in the development and location of signaling complexes. B7-2 predominantly enhances the overall response of Th2-type T cells, whereas B7-1 promotes the differentiation of Th1-type T cells (Suvas et al. 2002; Bhatia et al. 2005; Greenwald et al. 2005).

Role of Immune Checkpoints in Infectious Diseases

Owing to the widespread use of chronic infectious diseases in animals and the low efficiency of preventive measures, interest in studying animal immune checkpoints has increased. The study of cow immunity in leukemia has revealed an increase in the number of regulatory T cells responsible for the production of transforming growth factor- β (TGF- β). Natural killer (NK) cells, tumor necrosis factor (TNF- α), and interferon γ (IFN- γ) are all suppressed by the rise in TGF- β (Ohira et al. 2016). When studying BLV, a connection was established between the PD-1/PD-L1 receptor signaling pathway and lymphocyte activation gene 3 (LAG-3) (Okagawa et al. 2018). Additionally, elevated CTLA-4+ T cell expression has been reported during the progression of BLV infection (Suzuki et al. 2015). By enhancing the stimulation of macrophages and dendritic cells, T lymphocytes and their secretion of the cytokines IFN- γ and TNF- α play a significant role in developing immunity against bacterial and viral illnesses. However, in the late subclinical stages, T-lymphocyte activity decreases, contributing to an increased viral or

bacterial load and progression to clinical diseases (Sohal et al. 2008; Xing et al. 2022; Yang et al. 2023).

According to a recent study, tumors, malignancies, and persistent infections activate CTLA-4, which could compromise the immune system. Conversely, the immune system's defense against these illnesses is restored when antibodies prevent the action of the CTLA-4 receptor with CD80 or CD86 (Kaufmann et al. 2007). The inhibitory action of cattle CTLA-4 was proven in various investigations employing synthesized bovine CTLA-4-Ig. Anti-CTLA-4 antibodies were produced when mice were synthesized bovine CTLA-4-Ig. immunized with Antibodies against CTLA-4 protein significantly boosted both healthy and infected BLV immune system's IFN-γ production. According to the authors, antibodies against CTLA-4 may be useful for developing new therapies against BLV infections (Watari et al. 2019).

Similar effects were observed when the PD-1/PD1-L1 signal was blocked, which stimulates T-cell activation and proliferation in BLV. The progression of viral infection is aided by the association of PD-L1 on B cells, which lowers the number of PD-1+ T cells. Antibodies against PD-L1 or PD-1 were administered to HIV-infected macaques and mice infected with lymphocytic choriomeningitis virus (LCMV) to treat their infections. This restored multiple functions of the previously depleted T cells and eliminated the virus *in vivo*. According to studies, blocking the PD-1/PD-L1 pathway may have clinical uses for boosting host antimicrobial immunity for managing persistent infections (Ikebuchi et al. 2011).

The research of the impact of inhibiting the bovine PD-L1 receptor on the development of chronic ruminant enteritis caused by *Mycobacterium paratuberculosis* showed the possibility of anti-PD-L1 antibody treatment for regulating bacterial excretion. Treatment with anti-PD- L1 antibodies also activated the production of *M. paratuberculosis*-specific Th1 cytokines in infected cattle (Sajiki et al. 2021; Sun et al. 2021). However, immune checkpoint-based therapies for chronic infectious diseases in animals remain poorly understood (Sun et al. 2021).

Role of PD-1 in BLV

Subsets of the T cell family like Teff, CTL, Th, and Treg are essential for preventing viral illnesses and preserving internal homeostasis. Costimulatory or coinhibitory signaling pathways regulate T-cell subset functions following successful interaction of TCR with MHC. CD4+ T cells that specifically recognize MHC II molecules on Blymphocytes, macrophages, and NK cells are responsible for forming immunity against bacteria and parasites. CD8+ cells, which recognized MHC I after activation, function as cytotoxic cells in viral infections and malignancies.

However, T cells are depleted in chronic infections and malignant neoplasms owing to constant antigenic stimuli and inflammation. Lack of IL-2 and IFN-y and diminished cell proliferation prevent depleted T lymphocytes from performing their effector and cytotoxic activities (Wherry and Kurachi 2015; Dyck and Mills 2017). In cases of chronic infections and malignancies, T cells that are exhausted have been seen to show a higher level of induction of PD-1 and CTLA-4 receptors. (Bennett et al. 2003; Qin et al. 2019). Studies related to BLV have shown the inhibitory properties of PD-1 and CTLA-4 receptors during infection. Increased concentration receptors mRNA was observed in CD4+ and CD8+ cells of cows infected BLV. Depleted T cells also showed increased mRNA expression of factors such as LAG-3, T cell Ig and mucin domain-3 (TIM-3). In persistent infections and cancers, elevated PD-1 and TIM-3 mRNA levels aid in developing and maintaining pathogenic conditions. Simultaneously, antibodies blocking these signaling pathways reactivate Tcell depletion and activate immune responses (Ikebuchi et al. 2013; Nakamura et al. 2023).

Studies on depleted T cells from mice with chronic and acute infections caused by the LCMV showed different levels of PD-1 mRNA expression. Moreover, PD-1 blockade in T cells with different expression levels led to different results. PD-1 mRNA expression was significantly higher in mice with chronic viral infections. PD-1 blockade in mice with acute infection and high expression levels in cells did not restore the effector functions of depleted T cells (Yi et al. 2018). The provided data highlight the significant contribution of Teff proliferation to PD-1 blockage in improving the management of persistent viral infections. Moreover, the blocking of PD-1 and LAG-3 yielded good results, confirming the role of additional inhibitory in depleted T cells (Blackburn et al. 2009; Saeidi et al. 2018; Wykes and Lewin 2018).

When specific T lymphocytes are reduced, persistent infections in humans caused by the hepatitis B (HBV) and C (HCV) viruses and HIV also exhibit elevated expression of PD-1 on those cells. Moreover, the higher the level of PD-1 expression, the lower the activity of CD+ T cells (Dong et al. 2019). In BLV-infected cows with B-cell lymphoma, increased induction of PD-1 protein was observed in CD4+ T cells in the blood, and in CD4+ and CD8+ T cells in tumor-containing lymph nodes. Additionally, the number of PD-1+ T cells in lymph nodes

containing tumor was higher than that in the blood cells. Examination of other BLV lymph nodes from infected and healthy cows revealed low concentrations of PD+1 T cells. These data suggest that the tumor-bearing lymph nodes contain lymphoma-specific CD+ T cells. The PD-1 signaling pathway, however, enables BLV-induced lymphoma cells to prevent immune system reactions. (Ikebuchi et al. 2013).

In cattle, CD4+ T-cell growth and cytokine induction in response to viral infection are impaired in late-stage leukemia. PD-1 receptor blockade boosted IFN- γ induction in blood mononuclear cells in response to a mixture of glycoprotein gp51 peptides. The increase in IFN- γ was determined to be because of the increased PD-1+ cell levels in the CD4+ T cell population. Simultaneously, the blockade did not boost IL-10 production in mononuclear cells, indicating that PD-1+ T-cells function was not fully restored (Ikebuchi et al. 2013).

PD-1 inhibition is a viable method for reactivating fatigued T cells in BLV. Both cows with BLV infection and healthy cows produced more IFN- γ when exposed to monoclonal antibodies against bovine PD-1. Anti-PD-1 mAb therapy can be employed to treat various bovine infections despite the systemic side effects of immunotherapeutic techniques. Monoclonal antibodies may also expand education in the field of immunology and elucidate the immunosuppression disorder in chronic viral infection.

Role of CTLA-4 Protein in BLV

CTLA-4 is associated with T-cell depletion in several chronic infections. CTLA-4 is selectively induced in HIVspecific CD4+ T cells, whereas no induction of this antigen is in CD8+ T cells obtained from patients infected with HIV. Increased induction of CTLA-4 was observed in patients with HIV with advanced disease in which CD4+ T cells were unable to produce IL-2. CTLA-4 blockade enhances the functional activity of CD4 + T cells in vitro. These results demonstrate the potential role of CTLA-4 as a target for increasing CD4+ T-cell activity during immunotherapy in HIV-positive individuals (Kaufmann and Walker 2009; Wu et al. 2023). In CD4+CD25+ Foxp3+ T lymphocytes, CTLA-4 production is positively linked with disease progression. High levels of CTLA-4 have been observed in the leukocytes of HIV-infected macaques with a high viral load. The study demonstrated that CTLA-4 induction was lower in patients with HIV with slow disease progression than in asymptomatic patients with HIV. In addition, CTLA-4 induction was negatively correlated with CD4+ T-cell concentration, proving the significance of low Treg concentration combined with low CTLA-4 induction in slowing HIV progression (Boasso et al. 2007; Zhang et al. 2010).

Studies have shown that CTLA-4 contributes to the immunosuppression of chronic infections and malignancies. In persistent lymphocytosis in BLV-infected cows, T cell dysfunction leads to the dysregulation of Th1 and Th2 cytokines, resulting in disease progression. According to Suzuki et al. (2013), the disease progression in BLV-infected cows was closely related to the concentration of Foxp3+CD4+ T cells. These data suggest that decreased immunity during BLV infection is associated with the induction of CTLA-4 inhibitory molecules on Tregs. Amino acid sequencing of CTLA-4+ T cells revealed a conserved

MYPPPY site, which is characteristic of the CTLA-4 amino acid sequence in other mammals. Additionally, a protein phosphatase 2A binding site for the immunological inhibitory signaling pathway is present in a cytoplasmic domain of CTLA-4 (Suzuki et al. 2015).

In BLV infections, autoimmune diseases, graft rejection, persistent infection, and chronic viral infections, CTLA-4 is induced on CD4+CD25+Foxp3+T cells. Because Foxp3 is a marker of Tregs, studies support the assumption that CTLA-4 induces Tregs. The increase in Foxp3+CD4+cells level coincided with an increase in viral load, while there was a decrease in IFN- γ induction in BLV-infected cattle. These findings suggest that T cell depletion during BLV infection is associated with increased induction of CTLA-4 on Tregs (Suzuki et al. 2013).

The CTLA-4 inhibition has proven to be a successful treatment for malignant diseases and persistent infections, using melanoma and HIV as examples. Monoclonal antibodies against CTLA-4 activated T cells and cytokine production in BLV-infected cows. With T-cell activation, blocking CTLA-4 increased IFN-y production in BLV antigen-stimulated mononuclear cells of blood (Watari et al. 2022). The opposite effect on IFN-y increased concentration in mononuclear cells of blood activated by BLV antigens was exerted by bovine CTLA-4-Ig. The findings imply that CTLA-4 causes depletion of the function of CD4+ and CD8+ T cells and disease progression in cattle infected with BLV (Watari et al. 2019; Passariello et al. 2020; Lembo et al. 2022) and that antibodies against bovine CTLA-4 increase lymphocyte function and may be used as a novel treatment for chronic illnesses that have failed to respond to conventional therapies.

Conclusion

Owing to the widespread occurrence of BLV infections and limited studies on the immunosuppressive functions of Tregs, there is a scientific and technological need to obtain new knowledge and methodological approaches for treatment. In the onset and development of chronic infectious illnesses, immune checkpoint receptors and inhibitory cytokines mediate the immunoregulatory actions of Tregs. CTLA-4 and TGF-1 may facilitate t-cell-mediated immunosuppression during the persistent infection stage. The resulting recombinant proteins, monoclonal antibodies, and therapies for chronic infections can be used to identify novel targets and infections in farm animals.

PD-1, PD-L1, and CTLA-4 are potential targets to restore the function of exhausted T cells. Numerous studies using antibodies and peptide molecules to disrupt the PD-1 signaling cascade and cancer patient research studies support these results (Tao et al. 2020; Cao et al. 2023). Additionally, bovine checkpoint blockade studies have shown increased IFN- γ production in BLV-infected cattle. Based on the literature, the results contribute to research to develop new treatments for other types of infections in cattle. In addition, these results contribute to research in immunology, virology, bacteriology, and biotechnology aimed at elucidating the mechanisms of infectious diseases that cause immunosuppression.

Author Contributions

All authors contributed directly to the conception and design of the study, revision of the manuscript, and approval of the submitted version.

Acknowledgements

Funding: This research was funded by the Science Committee of the Ministry of Science and Education of the Republic of Kazakhstan (Grant No. AP09258581).

REFERENCES

- Agata Y, Kawasaki A, Nishimura H, Ishida Y, Tsubata T, Yagita H and Honjo T, 1996. Expression of the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. International Immunology 8: 765-772. <u>https://doi.org/10.1093/intimm/8.5.765</u>
- Attanasio J and Wherry EJ, 2016. Costimulatory and coinhibitory receptor pathways in infectious disease. Immunity 44: 1052-1068. https://doi.org/10.1016/j.immuni.2016.04.022
 Banchereau J and Palucka AK, 2005. Dendritic cells as
- Banchereau J and Palucka AK, 2005. Dendritic cells as therapeutic vaccines against cancer. Nature Review in Immunology 5: 296-306. <u>https://doi.org/10.1038/nri1592</u>
- Barber DL, Wherry EJ, Masopust D, Zhu B, Allison JP, Sharpe AH, Freeman GJ and Ahmed R, 2006. Restoring function in exhausted CD8 T cells during chronic viral infection. Nature 439: 682-687. <u>https://doi.org/10.1038/nature04444</u>
- Bennett F, Luxenberg D, Ling V, Wang IM, Marquette K, Lowe D, Khan N, Veldman G, Jacobs KA, Valge-Archer VE, Collins M and Carreno BM, 2003. Program death-1 engagement upon TCR activation has distinct effects on costimulation and cytokine-driven proliferation: attenuation of ICOS, IL-4, and IL-21, but not CD28, IL-7 and IL-15 responses. Journal of Immunology 170: 711-718. https://doi.org/10.4049/jimmunol.170.2.711
- Bhatia S, Edidin M, Almo SC and Nathenson SG, 2005. Different cell surface oligomeric states of B7-1 and B7-2: implications for signaling. Proceedings of National Academy of Sciences USA 102: 15569-15574. <u>https://doi.org/10.1073/pnas.</u> 0507257102
- Blackburn SD, Shin H, Haining WN, Zou T, Workman CJ, Polley A, Betts MR, Freeman GJ, Vignali DAA and Wherry EJ, 2009. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. Nature Immunology 10: 29-37. <u>https://doi.org/10.1038/ni.1679</u>
- Boasso A, Vaccari M, Hryniewicz A, Fuchs D, Nacsa J, Cecchinato V, Andersson A, Franchini G, Shearer GM and Chougnet C, 2007. Regulatory T-cell markers, indoleamine 2,3-dioxygenase, and virus levels in spleen and gut during progressive simian immunodeficiency virus infection. Journal of Virology 81: 11593-11603. <u>https://doi.org/</u> 10.1128/JVI.00760-07
- Buchbinder EI and Desai A, 2016. CTLA-4 and PD-1 Pathways: Similarities, Differences, and Implications of Their Inhibition. American Journal of Clinical Oncology 39: 98-106. https://doi.org/10.1097/COC.00000000000239
- Bucktrout SL, Bluestone JA and Ramsdell F, 2018. Recent advances in immunotherapies: from infection and autoimmunity, to cancer, and back again. Genome Medicine 10: 79. https://doi.org/10.1186/s13073-018-0588-4
- Buehring GC, DeLaney A, Shen H, Chu DL, Razavian N, Schwartz DA,Demkovich ZR and Bates MN, 2019. Bovine leukemia virus discovered in human blood. BMC Infectious Diseases 19: 297. https://doi.org/10.1186/s12879-019-3891-9
- Callahan MK, Wolchok JD and Allison JP, 2010. Anti-CTLA-4 antibody therapy: immune monitoring during clinical development of a novel immunotherapy. Semin Oncology 37: 473-484. <u>https://doi.org/10.1053/j.seminoncol.2010.</u> 09.001
- Canova R, Weber MN, Budaszewski RF, da Silva MS, Schwingel D, Canal CW and Kreutz LC, 2021. Bovine leukemia viral DNA found on human breast tissue is genetically related to the cattle virus. One Health 13: 100252. <u>https://doi.org/10.1016/j.onehlt.2021.100252</u>

- Cao H, Wu T, Zhou X, Xie S, Sun H, Sun Y and Li Y, 2023. Progress of research on PD-1/PD-L1 in leukemia. Frontiers in Immunology 14. <u>https://doi.org/10.3389/fimmu.2</u> 023.1265299
- Chen L, 2004. Co-inhibitory molecules of the B7-CD28 family in the control of T-cell immunity. Nature Reviews Immunology 4: 336-347. <u>https://doi.org/10.1038/nri1349</u>
- Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, Evdemon-Hogan M, Conejo-Garcia JR, Zhang L, Burow M, Zhu Y, Wei S, Kryczek I, Daniel B, Gordon A, Myers L, Lackner A, Disis ML, Knutson KL, Chen L and Zou W, 2004. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. Nature Medicine 10: 942-949. <u>https://doi.org/ 10.1038/nm1093</u>
- Danilovic DL, Mendes-Correa MC, Lima EU, Zambrini H, Barros KR and Marui S, 2012. Correlations of CTLA-4 gene polymorphisms and hepatitis C chronic infection. Liver International 32: 803-808. <u>https://doi.org/10.1111/j.1478-3231.2011.02694.x</u>
- de Quadros DL, Ribeiro VA, Rezende MA, Maté YA, Gomes MA, Secchi K, Strottmann DM, Frandoloso R and Kreutz LC, 2023. Oncogenic viral DNA related to human breast cancer found on cattle milk and meat. Comparative Immunology, Microbiology and Infectious Diseases 101. <u>https://doi.org/10.1016/j.cimid.2023.102053</u>
- do Nascimento AMM, de Souza CMS, Oliveira ACD, Blagitz MG, Ramos Sanchez EM, Della Libera AMMP, Leite RdMH, Fernandes ACdC and Souza FN, 2023. The bovine leukemia virus infection prolongs immunosuppression in dairy cows during the periparturient period by sustaining higher expression of immunological checkpoints in T cells. Veterinary Immunology and Immunopathology 263: 110636. https://doi.org/10.1016/j.vetimm.2023.110636
- Dong Y, Li X, Zhang L, Zhu Q, Chen C, Bao J and Chen Y, 2019. CD4+ T cell exhaustion revealed by high PD-1 and LAG-3 expression and the loss of helper T cell function in chronic hepatitis B. BMC Immunology 20: 27. <u>https://doi.org/</u> 10.1186/s12865-019-0309-9
- Dong Y, Sun Q and Zhang X, 2016. PD-1 and its ligands are important immune checkpoints in cancer. Oncotargets 8: 2171-2186. <u>https://doi.org/10.18632/oncotarget.13895</u>
- Dyck L and Mills KHG, 2017. Immune checkpoints and their inhibition in cancer and infectious diseases. European Journal of Immunology 47: 765-779. <u>https://doi.org/</u> 10.1002/eji.201646875
- Eskandari-Nasab E, Moghadampour M, Najibi H and Hadadi-Fishani M, 2014. Investigation of CTLA-4 and CD86 gene polymorphisms in Iranian patients with brucellosis infection. Microbiology & Immunology 58: 135-141. <u>https://doi.org/ 10.1111/1348-0421.12119</u>
- Farivar S, Dehghan Tezerjani M, Parvini N and Shiari R, 2014. Association of 1661A/G Cytotoxic T lymphocyte Antigen-4 (CTLA-4) Gene Polymorphism With a Clinical Subset of Iranian Children With Systemic Lupus Erythematosus. Thrita 3: e16020. <u>https://doi.org/10.5812/thrita.16020</u>
- Fife BT and Bluestone JA, 2008. Control of peripheral T-cell tolerance and autoimmunity via the CTLA-4 and PD-1 pathways. Immunological Reviews 224: 166-182. https://doi.org/10.1111/j.1600-065X.2008.00662.x
- Francisco LM, Sage PT and Sharpe AH, 2010. The PD-1 pathway in tolerance and autoimmunity. Immunology Reviews 236: 219-242. https://doi.org/10.1111/j.1600-065X.2010.00923.x
- Frauwirth KA, Riley JL, Harris MH, Parry RV, Rathmell JC, Plas DR, Elstrom RL, June CH and Thompson CB, 2002. The CD28 signaling pathway regulates glucose metabolism. Immunity 16: 769-777. <u>https://doi.org/10.1016/s1074-7613(02)00323-0</u>
- Freeman GJ, Gribben JG, Boussiotis VA, Ng JW, Restivo VA, Jr., Lombard LA, Gray GS and Nadler LM, 1993. Cloning

of B7-2: a CTLA-4 counter-receptor that costimulates human T cell proliferation. Science 262: 909-911. https://doi.org/10.1126/science.7694363

- Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, Fitz LJ, Malenkovich N, Okazaki T, Byrne MC, Horton HF, Fouser L, Carter L, Ling V, Bowman MR, Carreno BM, Collins M, Wood CR and Honjo T, 2000. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. Journal of Experimental Medicine 192: 1027-1034. <u>https://doi.org/10.1084/jem.192.7.1027</u>
- Gabrilovich D, 2004. Mechanisms and functional significance of tumour-induced dendritic-cell defects. Nature Reviews Immunology 4: 941-952. <u>https://doi.org/10.1038/nri1498</u>
- Greenwald RJ, Freeman GJ and Sharpe AH, 2005. The B7 family revisited. Annual Reviews in Immunology 23: 515-548. https://doi.org/10.1146/annurev.immunol.23.021704.11561 1
- Hathcock KS, Laszlo G, Dickler HB, Bradshaw J, Linsley P and Hodes RJ, 1993. Identification of an alternative CTLA-4 ligand costimulatory for T cell activation. Science 262: 905-907. <u>https://doi.org/10.1126/science.7694361</u>
- Homann D, Dummer W, Wolfe T, Rodrigo E, Theofilopoulos AN, Oldstone MB and Von Herrath MJ, 2006. Lack of intrinsic CTLA-4 expression has minimal effect on regulation of antiviral T-cell immunity. Journal of Virology 80: 270-280. https://doi.org/10.1128/JVI.80.1.270-280.2006
- Ikebuchi R, Konnai S, Okagawa T, Yokoyama K, Nakajima C, Suzuki Y, Murata S and Ohashi K, 2013. Blockade of bovine PD-1 increases T cell function and inhibits bovine leukemia virus expression in B cells in vitro. Veterinary Research 44: 59. https://doi.org/10.1186/1297-9716-44-59
- Ikebuchi R, Konnai S, Shirai T, Sunden Y, Murata S, Onuma M and Ohashi K, 2011. Increase of cells expressing PD-L1 in bovine leukemia virus infection and enhancement of antiviral immune responses in vitro via PD-L1 blockade. Veterinary Research 42: 103. <u>https://doi.org/10.1186/1297-9716-42-103</u>
- Ishida Y, Agata Y, Shibahara K and Honjo T, 1992. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. EMBO Journal 11: 3887-3895. <u>https://doi.org/10.1002/j.1460-2075.1992.tb05481.x</u>
- Jakubczik F, Jones K, Nichols J, Mansfield W, Cooke A and Holmes N, 2016. A SNP in the immunoregulatory molecule CTLA-4 controls mRNA Splicing in vivo but does not alter diabetes susceptibility in the NOD mouse. Diabetes 65: 120-128. https://doi.org/10.2337/db15-1175
- Jubel JM, Barbati ZR, Burger C, Wirtz DC and Schildberg FA, 2020. The role of PD-1 in acute and chronic infection. Frontiers in Immunology 11: 487. <u>https://doi.org/10.3389/ fimmu.2020.00487</u>
- Kao C, Oestreich KJ, Paley MA, Crawford A, Angelosanto JM, Ali MA, Intlekofer AM, Boss JM, Reiner SL, Weinmann AS and Wherry EJ, 2011. Transcription factor T-bet represses expression of the inhibitory receptor PD-1 and sustains virus-specific CD8+ T cell responses during chronic infection. Nature Immunology 12: 663-671. <u>https://doi.org/ 10.1038/ni.2046</u>
- Kaufmann DE, Kavanagh DG, Pereyra F, Zaunders JJ, Mackey EW, Miura T, Palmer S, Brockman M, Rathod A, Piechocka-Trocha A, Baker B, Zhu B, Le Gall S, Waring MT, Ahern R, Moss K, Kelleher AD, Coffin JM, Freeman GJ, Rosenberg ES and Walker BD, 2007. Upregulation of CTLA-4 by HIV-specific CD4+ T cells correlates with disease progression and defines a reversible immune dysfunction. Nature Immunology 8: 1246-1254. https://doi.org/10.1038/ni1515
- Kaufmann DE and Walker BD, 2009. PD-1 and CTLA-4 inhibitory cosignaling pathways in HIV infection and the

potential for therapeutic intervention. Journal of Immunology 182: 5891-5897. <u>https://doi.org/10.4049</u> /jimmunol.0803771

- Khatami A, Pormohammad A, Farzi R, Saadati H, Mehrabi M, Kiani SJ and Ghorbani S, 2020. Bovine Leukemia virus (BLV) and risk of breast cancer: a systematic review and meta-analysis of case-control studies. Infectious Agents of Cancer 15: 48. https://doi.org/10.1186/s13027-020-00314-7
- LaFleur MW, Muroyama Y, Drake CG and Sharpe AH, 2018. Inhibitors of the PD-1 Pathway in Tumor Therapy. Journal of Immunology 200: 375-383. <u>https://doi.org/10.4049/jimmunol.1701044</u>
- Latchman Y, Wood CR, Chernova T, Chaudhary D, Borde M, Chernova I, Iwai Y, Long AJ, Brown JA, Nunes R, Greenfield EA, Bourque K, Boussiotis VA, Carter LL, Carreno BM, Malenkovich N, Nishimura H, Okazaki T, Honjo T, Sharpe AH and Freeman GJ, 2001. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. Nature Immunology 2: 261-268. <u>https://doi.org/10.1038/85330</u>
- Lee KM, Chuang E, Griffin M, Khattri R, Hong DK, Zhang W, Straus D, Samelson LE, Thompson CB and Bluestone JA, 1998. Molecular basis of T cell inactivation by CTLA-4. Science 282: 2263-2266. <u>https://doi.org/10.1126/science. 282.5397.2263</u>
- Lembo RR, Manna L, Froechlich G, Sasso E, Passariello M and De Lorenzo C, 2022. New insights on the role of Anti-PD-L1 and Anti-CTLA-4 mAbs on different lymphocytes subpopulations in TNBC. Cancers (Basel) 14. https://doi.org/10.3390/cancers14215289
- Lindsten T, Lee KP, Harris ES, Petryniak B, Craighead N, Reynolds PJ, Lombard DB, Freeman GJ, Nadler LM and Gray GS, 1993. Characterization of CTLA-4 structure and expression on human T cells. Journal of Immunology 151: 3489-3499. PMID: 8397258
- Linsley PS, Greene JL, Brady W, Bajorath J, Ledbetter JA and Peach R, 1994. Human B7-1 (CD80) and B7-2 (CD86) bind with similar avidities but distinct kinetics to CD28 and CTLA-4 receptors. Immunity 1: 793-801. <u>https://doi.org/ 10.1016/s1074-7613(94)80021-9</u>
- Liu CP, Jiang JA, Wang T, Liu XM, Gao L, Zhu RR, Shen Y, Wu M, Xu T and Zhang XG, 2013. CTLA-4 and CD86 genetic variants and haplotypes in patients with rheumatoid arthritis in southeastern China. Genetics & Molecular Research 12: 1373-1382. https://doi.org/10.4238/2013.April.25.8
- Lorenz U, 2009. SHP-1 and SHP-2 in T cells: two phosphatases functioning at many levels. Immunology Reviews 228: 342-359. <u>https://doi.org/10.1111/j.1600-065X.2008.00760.x</u>
- Matsuzaki J, Gnjatic S, Mhawech-Fauceglia P, Beck A, Miller A, Tsuji T, Eppolito C, Qian F, Lele S, Shrikant P, Old LJ and Odunsi K, 2010. Tumor-infiltrating NY-ESO-1-specific CD8+ T cells are negatively regulated by LAG-3 and PD-1 in human ovarian cancer. Proceedings of National Acadademy of Sciences U S A 107: 7875-7880. https://doi.org/10.1073/pnas.1003345107
- Mayoux M, Roller A, Pulko V, Sammicheli S, Chen S, Sum E, Jost C, Fransen MF, Buser RB, Kowanetz M, Rommel K, Matos I, Colombetti S, Belousov A, Karanikas V, Ossendorp F, Hegde PS, Chen DS, Umana P, Perro M, Klein C and Xu W, 2020. Dendritic cells dictate responses to PD-L1 blockade cancer immunotherapy. Science Translational Medicine 12: eaav7431. <u>https://doi.org/10.1126/</u> scitranslmed.aav7431
- Mulley WR and Nikolic-Paterson DJ, 2008. Indoleamine 2,3dioxygenase in transplantation. Nephrology (Carlton) 13: 204-211. https://doi.org/10.1111/j.1440-1797.2007.00921.x
- Nakamoto N, Cho H, Shaked A, Olthoff K, Valiga ME, Kaminski M,Gostick E, Price DA, Freeman GJ, Wherry EJ and Chang KM, 2009. Synergistic reversal of intrahepatic HCV-specific CD8 T cell exhaustion by combined PD-1/CTLA-4

blockade. PLoS Pathogens 5: e1000313. <u>https://doi.org/</u> 10.1371/journal.ppat.1000313

- Nakamura H, Konnai S, Okagawa T, Maekawa N, Sajiki Y, Watari K, Kamitani K, Saito M, Kato Y, Suzuki Y, Shiro Murata and Ohashi K, 2023. Combined Immune checkpoint blockade enhances antiviral immunity against Bovine Leukemia Virus. Journal of Virology 97: e0143022. https://doi.org/10.1128/jvi.01430-22
- Oaks MK and Hallett KM, 2000. Cutting Edge: A soluble form of CTLA-4 in patients with autoimmune thyroid disease. The Journal of Immunology 164: 5015-5018. <u>https://doi.org/10.4049/jimmunol.164.10.5015</u>
- Ohira K, Nakahara A, Konnai S, Okagawa T, Nishimori A, Maekawa N, Ikebuchi R, Kohara J, Murata S and Ohashi K, 2016. Bovine leukemia virus reduces anti-viral cytokine activities and NK cytotoxicity by inducing TGF-β secretion from regulatory T cells. Immune Inflammatory Diseases 4: 52-63. https://doi.org/10.1002/iid3.93
- Okagawa T, Konnai S, Nishimori A, Maekawa N, Goto S, Ikebuchi R, Kohara J, Suzuki Y, Yamada S, Kato Y, Murata S and Ohashi K, 2018. Cooperation of PD-1 and LAG-3 in the exhaustion of CD4. Veterinary Research 49: 50. <u>https://doi.org/10.1186/s13567-018-0543-9</u>
- Parry RV, Chemnitz JM, Frauwirth KA, Lanfranco AR, Braunstein I, Kobayashi SV, Linsley PS, Thompson CR and Riley JL, 2005. CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. Molecular & Cellular Biology 25: 9543-9553. <u>https://doi.org/10.1128/MCB.</u> 25.21.9543-9553.2005
- Passariello M, Vetrei C, Sasso E, Froechlich G, Gentile C, D'alise AM, Zambrano N, Scarselli E, Nicosia A and De Lorenzo C, 2020. Isolation of two novel human anti-ctla-4 mabs with intriguing biological properties on tumor and nk cells. Cancers (Basel) 12: 1-24. <u>https://doi.org/10.3390/cancers</u> 12082204
- Qin W, Hu L, Zhang X, Jiang S, Li J, Zhang Z and Wang X, 2019. The Diverse Function of PD-1/PD-L Pathway Beyond Cancer. Frontiers in Immunology 10: 2298. <u>https://doi.org/ 10.3389/fimmu.2019.02298</u>
- Rochmah N, Faizi M, Nova S, Setyoningrum RA, Basuki S and Endaryanto A, 2022. CTLA-4 CT-60 A/G and CTLA-4 1822 C/T gene polymorphisms in Indonesians with Type 1 diabetes mellitus. Applications in Clinical Genetics 15: 19-25. https://doi.org/10.2147/TACG.S359158
- Rudd CE, Taylor A and Schneider H, 2009. CD28 and CTLA-4 coreceptor expression and signal transduction. Immunology Reviews 229: 12-26. <u>https://doi.org/10.1111/j.1600-065X.2009.00770.x</u>
- Rui M, Zhang W, Mi K, Ni H, Ji W, Yu X, Qin J and Feng C, 2023. Design and evaluation of α-helix-based peptide inhibitors for blocking PD-1/PD-L1 interaction. International Journal of Biological Macromolecules 253. <u>https://doi.org/10.1016/j.ijbiomac.2023.126811</u>
- Saeidi A, Zandi K, Cheok YY, Saeidi H, Wong WF, Lee CYQ, Cheong HC, Yong YK, Larsson M and Shankar EM, 2018. T-Cell exhaustion in chronic infections: Reversing the state of exhaustion and reinvigorating optimal protective immune responses. Frontiers in Immunology 9: 2569. <u>https://doi.org/ 10.3389/fimmu.2018.02569</u>
- Saito H, Kuroda H, Matsunaga T, Osaki T and Ikeguchi M, 2013. Increased PD-1 expression on CD4+ and CD8+ T cells is involved in immune evasion in gastric cancer. Journal of Surgical Oncology 107: 517-522. <u>https://doi.org/10.1002/ jso.23281</u>
- Sajiki Y, Konnai S, Nagata R, Kawaji S, Nakamura H, Fujisawa S, Okagawa T, Maekawa N, Kato Y, Suzuki Y, Murata S, Mori Y and Ohashi K, 2021. The enhancement of Th1 immune response by anti-PD-L1 antibody in cattle infected with Mycobacterium avium subsp. paratuberculosis. The

Journal of Veterinary Medical Science 83: 162-166. https://doi.org/10.1292/jvms.20-0590

- Sanmamed MF and Chen L, 2014. Inducible expression of B7-H1 (PD-L1) and its selective role in tumor site immune modulation. Cancer Journal 20: 256-261. <u>https://doi.org/10.1097/PPO.000000000000061</u>
- Schneider H, Downey J, Smith A, Zinselmeyer BH, Rush C, Brewer JM, Brewer JM, Wei B, Hogg N, Garside P and Rudd CE, 2006. Reversal of the TCR stop signal by CTLA-4. Science 313: 1972-1975. <u>https://doi.org/10.1126/science. 1131078</u>
- Sharpe AH and Pauken KE, 2018. The diverse functions of the PD1 inhibitory pathway. Nature Reviews in Immunology 18: 153-167. <u>https://doi.org/10.1038/nri.2017.108</u>
- Shen X and Zhao B, 2018. Efficacy of PD-1 or PD-L1 inhibitors and PD-L1 expression status in cancer: meta-analysis. BMJ 362: k3529. https://doi.org/10.1136/bmj.k3529
- Shinohara T, Taniwaki M, Ishida Y, Kawaichi M and Honjo T, 1994. Structure and chromosomal localization of the human PD-1 gene (PDCD1). Genomics 23: 704-706. <u>https://doi.org/ 10.1006/geno.1994.1562</u>
- Shirai T, Konnai S, Ikebuchi R, Okagawa T, Suzuki S, Sunden Y, Onuma M, Murata S and Ohashi K, 2011. Molecular cloning of bovine lymphocyte activation gene-3 and its expression characteristics in bovine leukemia virus-infected cattle. Veterinary Immunology & Immunopathology 144: 462-467. <u>https://doi.org/10.1016/j.vetimm.2011.08.018</u>
- Sohal JS, Singh SV, Tyagi P, Subhodh S, Singh PK, Singh AV, Narayanasamy K, Sheoran N and Sandhu KS, 2008. Immunology of mycobacterial infections: with special reference to Mycobacterium avium subspecies paratuberculosis. Immunobiology 213: 585-598. <u>https://doi.org/10.1016/j.imbio.2007.11.002</u>
- Starr R, Willson TA, Viney EM, Murray LJ, Rayner JR, Jenkins BJ, Gonda TJ, Alexander WS, Metcalf D, Nicola NA and Hilton DJ, 1997. A family of cytokine-inducible inhibitors of signalling. Nature 387: 917-921. <u>https://doi.org/ 10.1038/43206</u>
- Sun HL, Du XF, Tang YX, Li GQ, Yang SY, Wang LH, Li XW, Ma CJ and Jiang RM, 2021. Impact of immune checkpoint molecules on FoxP3+ Treg cells and related cytokines in patients with acute and chronic brucellosis. BMC Infectious Diseases 21: 1025. <u>https://doi.org/10.1186/s12879-021-06730-3</u>
- Sutherland RM, Brady JL, Georgiou HM, Thomas HE and Lew AM, 2000. Protective effect of CTLA4Ig secreted by transgenic fetal pancreas allografts. Transplantation 69: 1806-1812. https://doi.org/10.1097/00007890-200005150-00013
- Suvas S, Singh V, Sahdev S, Vohra H and Agrewala JN, 2002. Distinct role of CD80 and CD86 in the regulation of the activation of B Cell and B Cell Lymphoma. Journal of Biological Chemistry 277: 7766-7775. <u>https://doi.org/ 10.1074/jbc.M105902200</u>
- Suzuki S, Konnai S, Okagawa T, Ikebuchi R, Nishimori A, Kohara J, Mingala CN, Murata S and Ohashi K, 2015. Increased expression of the regulatory T cell-associated marker CTLA-4 in bovine leukemia virus infection. Veterinary Immunology & Immunopathology 163: 115-124. <u>https://doi.org/10.1016/j.vetimm.2014.10.006</u>
- Suzuki S, Konnai S, Okagawa T, Ikebuchi R, Shirai T, Sunden Y, Mingala CN, Murata S and Ohashi K, 2013. Expression analysis of Foxp3 in T cells from bovine leukemia virus

infected cattle. Microbiology and Immunology 57: 600-604. https://doi.org/10.1111/1348-0421.12073

- Tao H, Cheng L, Liu L, Wang H, Jiang Z, Qiang X, Xing L, Xu Y, Cai X, Yao J, Wang M and Qiu Z, 2020. A PD-1 peptide antagonist exhibits potent anti-tumor and immune regulatory activity. Cancer Letters 493: 91-101. <u>https://doi.org/ 10.1016/j.canlet.2020.08.009</u>
- Watari K, Konnai S, Maekawa N, Okagawa T, Suzuki Y, Murata S and Ohashi K, 2019. Immune inhibitory function of bovine CTLA-4 and the effects of its blockade in IFN-γ production. BMC Veterinary Research 15: 380. <u>https://doi.org/10.1186/ s12917-019-2082-7</u>
- Watari K, Konnai S, Okagawa T, Maekawa N, Sajiki Y, Kato Y, Suzuki Y, Murata S and Ohashi K, 2022. Enhancement of interleukin-2 production by bovine peripheral blood mononuclear cells treated with the combination of antiprogrammed death-ligand 1 and cytotoxic T lymphocyte antigen 4 chimeric monoclonal antibodies. Journal of Veterinary Medical Sciences 84: 6-15. <u>https://doi.org/ 10.1292/jvms.21-0552</u>
- Wherry EJ and Kurachi M, 2015. Molecular and cellular insights into T cell exhaustion. Nature Reviews Immunology 15: 486-499. https://doi.org/10.1038/nri3862
- Wing K, Yamaguchi T and Sakaguchi S, 2011. Cell-autonomous and -non-autonomous roles of CTLA-4 in immune regulation. Trends in Immunology 32: 428-433. <u>https://doi.org/10.1016/j.it.2011.06.002</u>
- Wu L, Su J, Yang J, Gu L, Zhang R, Liu L, Lu H and Chen J, 2023. Use of programmed cell death protein 1 (PD-1) inhibitor therapy in HIV-infected patients with advanced cancer: a single-center study from China. Infectious Agents in Cancer 18: 35. <u>https://doi.org/10.1186/s13027-023-00512-z</u>
- Wykes MN and Lewin SR, 2018. Immune checkpoint blockade in infectious diseases. Nature Reviews Immunology 18: 91-104. https://doi.org/10.1038/nri.2017.112
- Xing K, Zhou P, Li J, Liu M and Zhang WE, 2022. Inhibitory effect of PD-1/PD-L1 and blockade immunotherapy in leukemia. Combinatorial Chemistry and High Throughput Screening 25: 1399-1410. <u>https://doi.org/10.2174/157</u> <u>489361666621 0707101516</u>
- Yang S, Wang Y, Yu F, Cheng R, Zhang Y, Zhou D, Ren X, Deng Z and Zhao H, 2023. Structural and functional insights into the modulation of T cell costimulation by monkeypox virus protein M2. Nature Communications 14. <u>https://doi.org/ 10.1038/s41467-023-40748-2</u>
- Yi M, Jiao D, Xu H, Liu Q, Zhao W, Han X and Wu K, 2018. Biomarkers for predicting efficacy of PD-1/PD-L1 inhibitors. Molecular Cancer 17: 129. <u>https://doi.org/ 10.1186/s12943-018-0864-3</u>
- Zhang XF, Pan K, Weng DS, Chen CL, Wang QJ, Zhao JJ, Pan QZ, Liu Q, Jiang SS, Li YQ, Zhang HX and Xia JC, 2016. Cytotoxic T lymphocyte antigen-4 expression in esophageal carcinoma: implications for prognosis. Oncotargets 7: 26670-26679. <u>https://doi.org/10.18632/oncotarget.8476</u>
- Zhang Z, Jiang Y, Zhang M, Liu J, Sun G, Shi W, Wang Y and Shang H, 2010. Alterations of CD4(+) CD25(+) Foxp3(+) regulatory T cells in HIV-infected slow progressors of former blood donors in China. Microbiology & Immunology 54: 625-633. <u>https://doi.org/10.1111/j.1348-0421.2010.</u> 00259.x
- Zou W and Chen L, 2008. Inhibitory B7-family molecules in the tumour microenvironment. Nature Reviews Immunology 8: 467-477. <u>https://doi.org/10.1038/nri2326</u>